

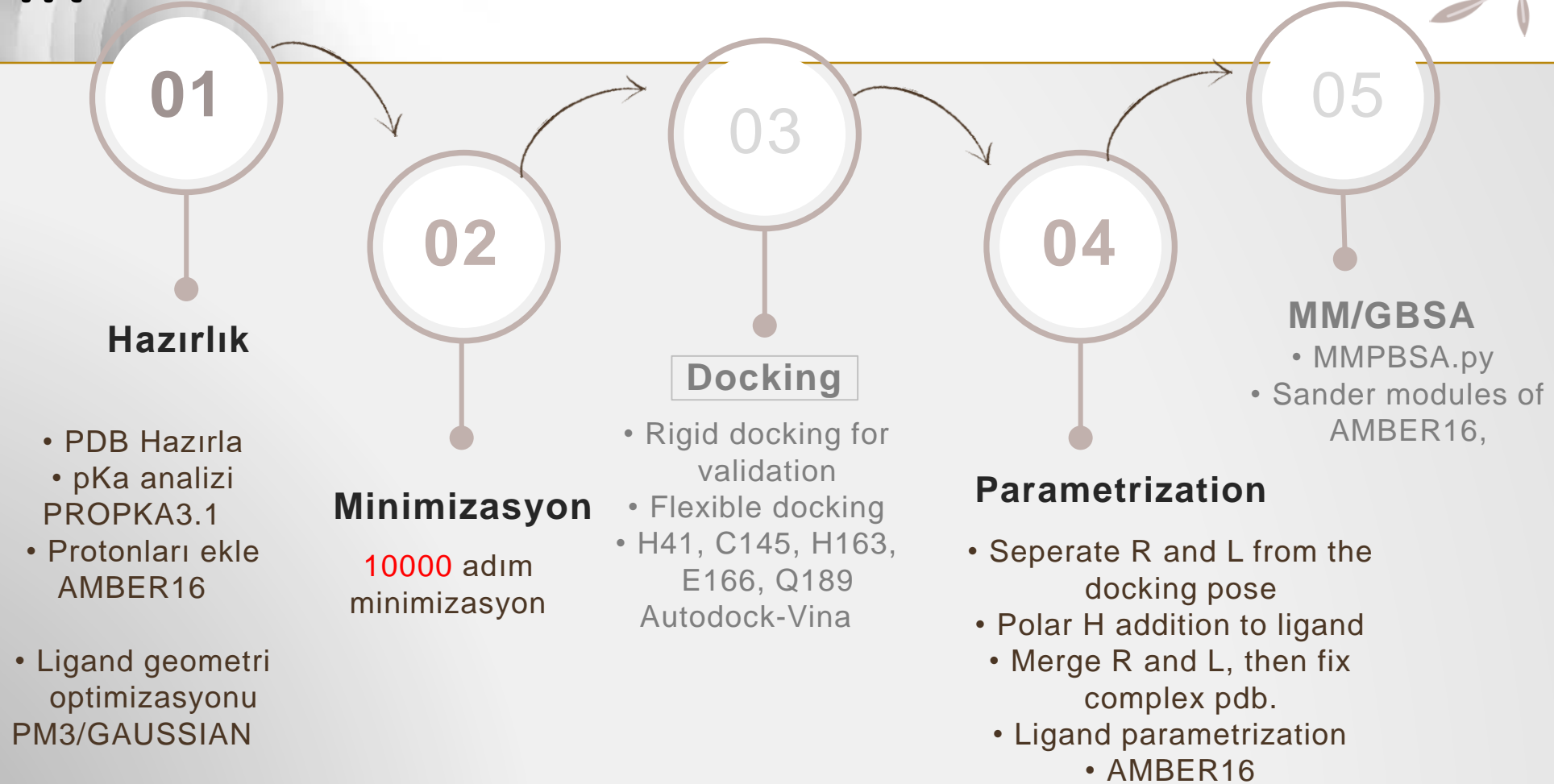
EURO

Moleküler Dinamik Simülasyonların ABC'si

30 Eylül 2022

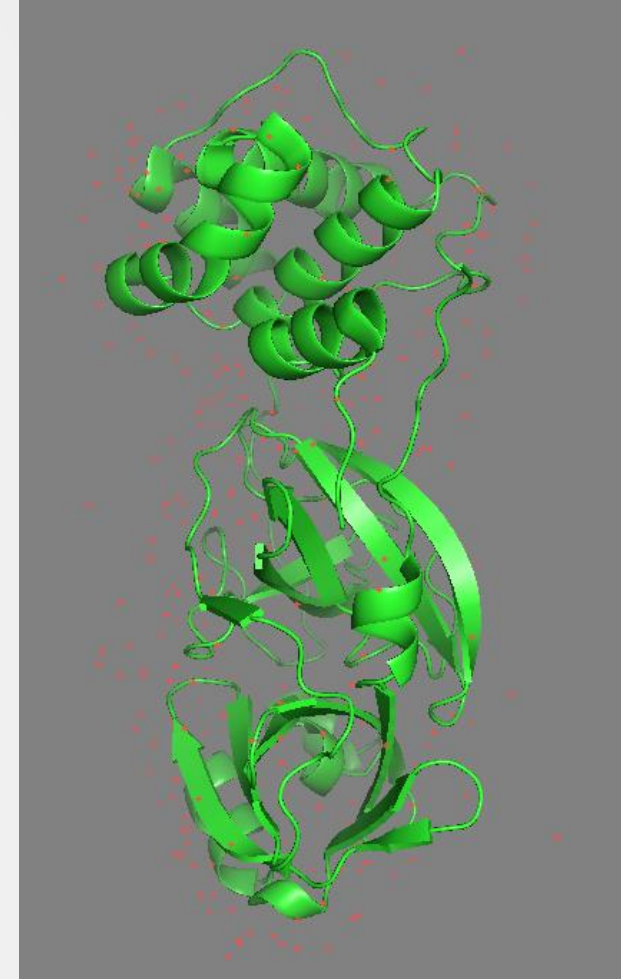
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İçerik



-Enzim yapısı hakkında bilgi toplama

- Main protease (Mpro, 3CLpro)
- Katalitik aktivite için dimer zorunlu
- Başka türlü S1 cebi oluşmuyor ve bağlanma gerçekleşmiyor.
- His-41 ve Cys-145 aktif bölge rezidüleri
- Sekans uzunluğu : 306 a.a
- Pdb yapısı: 6lu7.pdb



- Pymol betiđi (pdbholo-prep.pml dosyası)
- 1. X-ray yapısını yükle (.cif ya da pdb formatında-yapıyı saklayan pymol obje'ye aktar)
- 2.Suları kaldır
- 3.Monomerleri ayır
- 4.Bütünü saklayan objeyi sil
- 5.Zincirleri isimlendir
- 6.Yeniden yapılandır
- 7.Yapı oluşan dimer yapıyı kaydet

Bu aşamada ligand ve diğer hetero yapılar Pymol ile silinir. 6lu7-dimer.pdb hazır hale gelir.

- `pdb4amber -i 6lu7-dimer.pdb -o 6lu7-fixed.pdb`

```
=====
Summary of pdb4amber for file 6lu7-dimer.pdb
=====
```

```
-----Chains
```

```
The following (original) chains have been found:
```

```
A
B
```

```
----- Histidines (Renumbered Residues!)
```

```
It was not possible to determine the protonation state of the following HIS
residues based on presence of hydrogens. Amber will consider them as HIE
(epsilon-HIS) by default. If other protonation state desired change to HID
(delta-HIS) or HIP (protonated HIS) by hand.
```

```
HIS_64
HIS_386
HIS_163
HIS_164
HIS_470
HIS_552
HIS_41
HIS_172
HIS_80
HIS_370
HIS_469
HIS_246
HIS_347
HIS_478
```

```
----- Cysteines in Disulfide Bonds (Renumbered Residues!)
```

```
No disulfide bonds have been detected.
```

```
----- Missing Heavy Atoms (Renumbered Residues!)
```

```
None
```

- Pdb4amber ön kontrolü yaparak protein yapısındaki hataları temizler **ANCAK**
- Eksik rezidü var ise HOMOLOGU MODELLING yapılmalı.
- Protonasyon durumlarına bir sonraki adımda karar verilmeli

- <https://ambermd.org/tutorials/basic/tutorial9/index.php>

- Hesaplanan **pKa**
>>>>**pH**_{ortam} yan zincir protonlanır. .pdb dosyası elde edilen sonuçlar ve yapılan gözlemler sonucunda editlenir.
- pKa değerleri with PROPKA3 ve H++ web server (v3.2) ile deneyin belirttiği pH değerinde hesaplanır.
- Eğer kompleks (reseptör-ligant) yapınız var ise hesaplar için **MUTLAKA** Propka3.1 kullanmanız gerekir.

- **Histidine**

HID: Histidine with hydrogen on the delta nitrogen

HIE: Histidine with hydrogen on the epsilon nitrogen

HIP: Histidine with hydrogens on both nitrogens; this is positively charged.

- **Lysine**

LYS protonated lysine (+1 total charge)

LYN deprotonated (neutral) lysine

- **Cysteine**

CYM: deprotonated cysteine

CYX: Disulfide bridge

CYS: protonated cysteine

- **Aspartic acid**

ASP - deprotonated aspartate

ASH - protonated aspartic acid (nl)

GLU - deprotonated histidine

GLH - protonated histidine (nl)

Residue@Chain	6lu7 (Complex)	
	H++	PROPKA3.1
HIS-41@A, HIS-347@B	0.28	4.29
HIS-64@A, HIS-370@B	6.67	6.22
HIS-80@A, HIS-386@B	6.60	5.77
HIS-163@A, HIS-469@B	<0.0	1.37 (1.05)
HIS-164@A, HIS-470@B	2.80	1.86
HIS-172@A, HIS-478@B	3.84	6.47 (5.41)
HIS-246@A, HIS-552@B	6.03	5.43

Residue@Chain	Options	Proposed
HIS-41@A, HIS-347@B	HID	HID
HIS-64@A, HIS-370@B	HID/E/P	HIP
HIS-80@A, HIS-386@B	HIE/D/P	HIP
HIS-163@A, HIS-469@B	HIE	HIE
HIS-164@A, HIS-470@B	HID/E	HID
HIS-172@A, HIS-478@B	HIE/D	HIE
HIS-246@A, HIS-552@B	HIE/D/P	HIP

Asp, Glu, Cys ve Lys da kontrol edilir. Onlar için önerilenler uygun bulundu ve sadece HIS leri değiştirmek üzere devam ediyoruz.


```
/okyanus/users/fsungur/EUROCC_WORKSHOP/HOLO-6LU7
```

Rezidüleri tekrar adlandırmak için (relabel-HISTIDINE)

```
cp 6lu7-fixed_renum.txt 6lu7-fixed_renum-modify.txt
```

```
cat 6lu7-fixed_renum-modify.txt | grep HI | \awk '{ printf "s/%3s A %3s /%3s A %3s /\n", $1, $4, $3, $4}' > sed.in
```

UNUTMA:

- sed.in dosyasında düzenlemeler sonrasında

```
sed -f sed.in 6lu7-fixed.pdb > 6lu7-in.pdb
```

6lu7-in.pdb dosyasında tüm histidinler uygun protonasyon durumlarıyla tanımlanmıştır.
Diğer polar rezidüler için önerilenlerden başka bir değişiklik yapılmayacak.

tleap nasıl etkin kullanılır?

<http://ambermd.org/tutorials/pengfei/>

https://wikis.ch.cam.ac.uk/ro-walesdocs/wiki/index.php/Simple_scripts_for_LEaP_to_create_topology_and_coordinate_files

```
# load the force field libraries
source leaprc.protein.ff14SB
source leaprc.water.tip3p

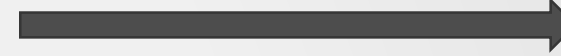
# load the PDB file , create a "mol" object
mol = loadpdb 6lu7-in.pdb

# neutralize system
addions mol Na+ 0
addions mol Cl - 0

# save topology and coordinate files

saveamberparm mol 6lu7-gas.top 6lu7-gas.incrd
quit
```

ÇALIŞTIR
tleap -f leap
.in



ÇIKTI DOSYALAR
6lu7-gas.top
6lu7-gas.incrd

Oluşan yapının görseli

```
ambpdb -p 6lu7-gas.top < 6lu7-gas.incrd > 6lu7-  
gas.pdb
```

Sistem Hazırlama-Minimizasyon minH.in (sadece hidrojenler)

/okyanus/users/fsungur/EUROCC_WORKSHOP/HOLO-6LU7/Hminimization/short

minH.in

MD 10K NVT strong coupling, restraints on all protein heavy atoms

&cntrl

imin = 1, !minimize initial structure

ntx = 1, irest=0, ! read coordinate

ntmin = 1, !both SD and CG is going to be used if =1 only SD no need for nyc

ncyc=500, !switch from steepest descent to CG after 1000 cycles

maxcyc = 1000, !max cycle for minimization

ntpr=10, !print to mdout every ntpr steps

ntb=0, !non periodic simulation

ntp=0, !no pressure scaling

cut = 9999., !Nonbonded cutoff in Angstrom

rgbmax=15., !controls the maximum distance between atom pairs

ntr = 1, restraint_wt = 10.0, restraintmask = '!@H='

/

ambpdb -p 6lu7-gas.top -c 6lu7-gas-min0.rst > 6lu7-gas-min0.pdb

```
#!/bin/bash
```

```
#SBATCH -J "Tutorial"           # isin adi
```

```
#SBATCH -A cmpei1              # account / proje adi
```

```
#SBATCH -p shortq              # kuyruk (partition/queue) adi
```

```
#SBATCH -n 8
```

```
#SBATCH -N 1                    # cekirdek / islemci sayisi
```

```
module load cuda/cuda-8.0
```

```
module load intel/parallel_studio_xe_2017.0.035
```

```
module load intel/impi_2017.0.098
```

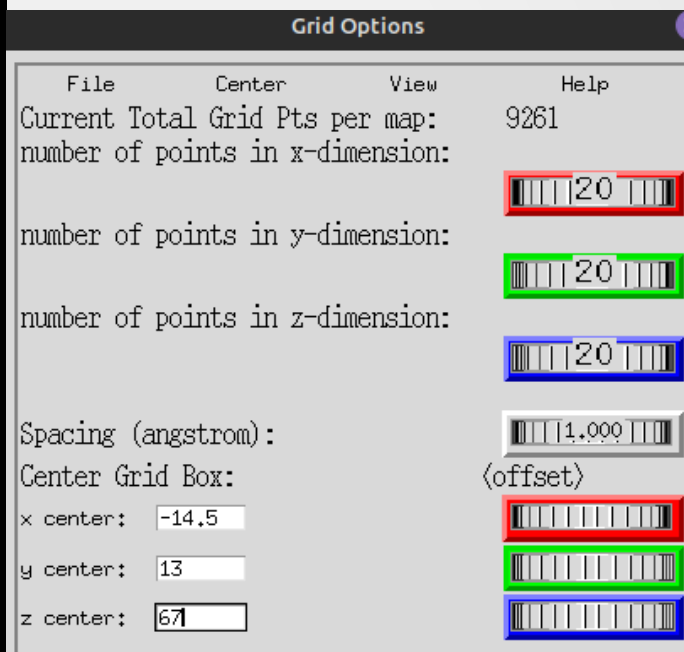
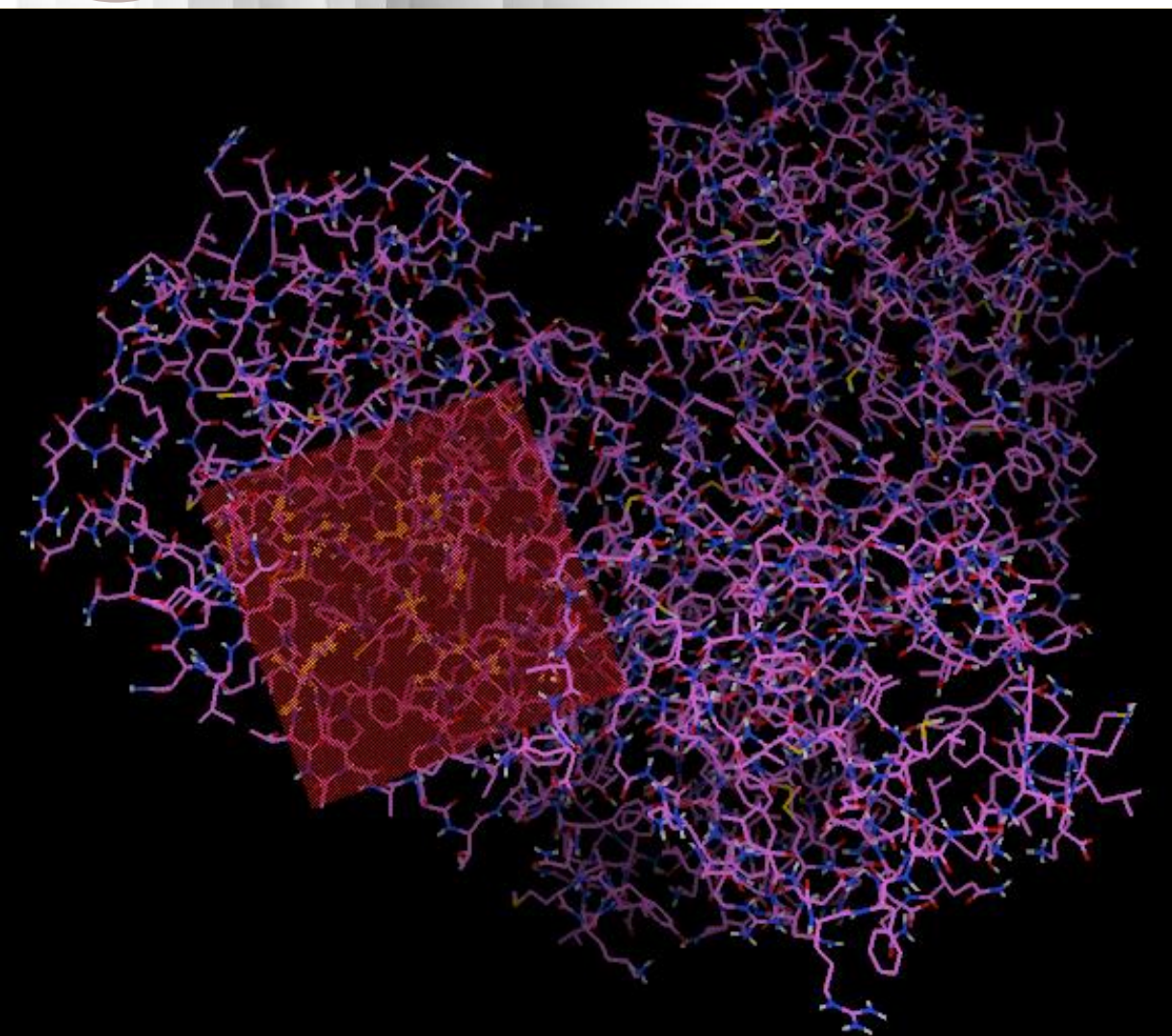
```
module load intel/mkl_2017.0.098
```

```
source /okyanus/progs/amber/amber16/intel/amber.sh
```

```
#calisacak is
```

```
$AMBERHOME/bin/sander -O -i minH.in -o 6lu7-gas-min0.out -p 6lu7-gas.top -c 6lu7-gas.incrd -ref 6lu7-gas.incrd  
-r 6lu7-gas-min0.rst -e 6lu7-gas-min0.ene -v 6lu7-gas-min0.vel -x 6lu7-gas-min0.incrd -inf 6lu7min.mdinfo
```

1. Minimize edilen protein yapısı, kenetleme için hazırlanır.
2. Esnek kenetleme çalışması için, proteinin sabit ve esnek kısımları ayrı PDB dosyalarına ayrılır. Bu aşamada hareket etmesine izin verilen yani esnek bırakılan aminoasitlerin hareket edebilmelerini sağlayan dönebilir bağlar belirlenir.
3. Kenetlenecek ligand hazırlanır. Ligandın aktif bölge içerisinde hareket ederek uygun lokasyonda konumlanması için esnek bırakılır ve dönebilir bağları belirlenir.
4. Kenetleme için gerekli parametrelerin yer aldığı konfigürasyon dosyası hazırlanır ve iş başlatılır.



Konfigürasyon dosyası
(conf.txt)

```

1 receptor=6lu7-prep_rigid.pdbqt
2 flex=6lu7-prep_flex.pdbqt
3 ligand=ligand.pdbqt
4
5 center_x = -14.5
6 center_y = 13
7 center_z = 67
8
9 size_x = 20
10 size_y = 20
11 size_z = 20
12
13 exhaustiveness = 32
14
15 out=ligand_out.pdbqt
16 log=ligand_log.txt

```



```
1 #!/bin/bash
2
3 #SBATCH -J "Tutorial"           # isin adi
4 #SBATCH -A cmpeil              # account / proje adi
5 #SBATCH -p defq                # kuyruk (partition/queue) adi
6 #SBATCH -n 28
7 #SBATCH -N 1                   # cekirdek / islemci sayisi
8
9 mgltools_scripts_path=/okyanus/users/fsungur/mgltools_x86_64Linux2_1.5.7/MGLToolsPckgs/AutoDockTools/Utilities24
10 vina_path=/okyanus/users/fsungur/autodock_vina_1_1_2_linux_x86/bin
11 pythonsh=/okyanus/users/fsungur/mgltools_x86_64Linux2_1.5.7/bin/pythonsh
12
13 # 1. Reseptör hazırlanması
14 $pythonsh $mgltools_scripts_path/prepare_receptor4.py -r 6lu7-gas-min0.pdb -o 6lu7-prep.pdbqt
15
16 # 2. Reseptörün sabit ve esnek kısımlarının hazırlanması
17 $pythonsh $mgltools_scripts_path/prepare_flexreceptor4.py -r 6lu7-prep.pdbqt -s HID41_CYS145_HIE163_GLU166_GLN189
18
19 # 3. Ligandın hazırlanması
20 $pythonsh $mgltools_scripts_path/prepare_ligand4.py -l ligand.pdb
21
22 # 4. Kenetleme
23 $vina_path/vina --config conf.txt
```

input files

- ligand.pdb
- receptor.pdb

Pymol kullanarak complex.pdb adıyla kayıt edebilirsiniz. 306 ve 307 arasına TER keyword eklemek gerekiyor.

Then use *pd4amber* to fix pdb file.

- ```
> pdb4amber -i ligand.pdb -o ligand-clean.pdb
> pdb4amber -i complex.pdb -o complex-clean.pdb
```

Use antechamber program to parametrize ligand-clean.pdb

```
> antechamber -i ligand-clean.pdb -fi pdb -at gaff -o ligand.mol2 -fo mol2 -c bcc -s 2 -j 5
```

Use parmchk2 utility to test and generate parameter files

```
> parmchk2 -s 2 -i ligand.mol2 -f mol2 -o ligand.frcmod
```

```
> parmchk2 -s 2 -i ligand.mol2 -f mol2 -o ligand.frcmod.full -a Y
```

**Check ligand.frcmod file: You should check these parameters carefully before running a simulation. If antechamber can't empirically calculate a value or has no analogy it will either add a default value that it thinks is reasonable or alternatively insert a place holder (with zeros everywhere) and the comment 'ATTN needs revision'. In this case you will have to manually parameterise this yourself.**

# Antechamber



- This is the most important program in the package. It can perform many file conversions, and can also assign atomic charges and atom types.
  - -c : charge method
  - -j : atom type and bond type prediction index, default is 4
    - 0 : no assignment
    - 1 : atom type
    - 2 : full bond types
    - 3 : part bond types
    - 4 : atom and full bond type
    - 5 : atom and part bond type

## List of the Charge Methods:

| charge method    | abbre. | index | charge method  | abbre. | index |
|------------------|--------|-------|----------------|--------|-------|
| RESP             | resp   | 1     | AM1-BCC        | bcc    | 2     |
| CM1              | cm1    | 3     | CM2            | cm2    | 4     |
| ESP (Kollman)    | esp    | 5     | Mulliken       | mul    | 6     |
| Gasteiger        | gas    | 7     | Read in charge | rc     | 8     |
| Write out charge | wc     | 9     | Delete Charge  | dc     | 10    |

## List of the File Formats:

| file format       | type | abbre. | index | file format        | type | abbre. | index |
|-------------------|------|--------|-------|--------------------|------|--------|-------|
| Antechamber       |      | ac     | 1     | Sybyl Mol2         |      | mol2   | 2     |
| PDB               |      | pdb    | 3     | Modified PDB       |      | mpdb   | 4     |
| AMBER PREP (int)  |      | prepi  | 5     | AMBER PREP (car)   |      | prepc  | 6     |
| Gaussian Z-Matrix |      | gzmat  | 7     | Gaussian Cartesian |      | gcrt   | 8     |
| Mopac Internal    |      | mopint | 9     | Mopac Cartesian    |      | mopcrt | 10    |
| Gaussian Output   |      | gout   | 11    | Mopac Output       |      | mopout | 12    |
| Alchemy           |      | alc    | 13    | CSD                |      | csd    | 14    |
| MDL               |      | mdl    | 15    | Hyper              |      | hin    | 16    |
| AMBER Restart     |      | rst    | 17    | Jaguar Cartesian   |      | jcrt   | 18    |
| Jaguar Z-Matrix   |      | jzmat  | 19    | Jaguar Output      |      | jout   | 20    |
| Divcon Input      |      | divcrt | 21    | Divcon Output      |      | divout | 22    |
| SQM Input         |      | sqmcr  | 23    | SQM Output         |      | sqmout | 24    |
| Charmm            |      | charmm | 25    | Gaussian ESP       |      | gesp   | 26    |

Use tleap to generate parameter library file for ligand

**input files**

– **ligand.mol2**

> tleap -f leap-ligand .in

```
source leaprc.protein.ff14SB
source leaprc.gaff

LIG = loadmol2 ligand.mol2

check LIG

loadamberparams ligand.frcmod

check LIG

saveoff LIG ligand.lib

quit
```

Use tleap to generate topology and coordinate files for MD simulation

## input files

- ligand.lib
- ligand.frcmod
- ligand.mol2
- receptor-clean.pdb
- complex-clean.pdb

> tleap -f leap-complex.in

Generate complex-solvated.pdb

```
source leaprc.protein.ff14SB
source leaprc.water.tip3p
source leaprc.gaff

loadamberparams ligand.frcmod
loadoff ligand.lib

COMPLEX = loadpdb complex-clean.pdb
LIGAND = loadmol2 ligand.mol2
RECEPTOR = loadpdb receptor-clean.pdb

set default PBRadii mbondi2

saveamberparm COMPLEX receptor-ligand.prmtop receptor-ligand.incrd

addions COMPLEX Na+ 0
addions COMPLEX Cl- 0

saveamberparm COMPLEX complex.prmtop complex.incrd
saveamberparm LIGAND ligand.prmtop ligand.incrd
saveamberparm RECEPTOR receptor.prmtop receptor.incrd

solvatebox COMPLEX TIP3PBOX 12.0 iso 1.00
set COMPLEX box 111.75708

saveamberparm COMPLEX complex-solvated.prmtop complex-solvated.incrd

quit
```

> ambpdb -p complex-solvated.prmtop -c complex-solvated.incrd > complex-solvated.pdb

# Thanks!



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